

Concise Synthesis of (S)-N-BOC-2-Hydroxymethylmorpholine and (S)-N-BOC-Morpholine-2-carboxylic Acid

Kevin E. Henegar*

Pfizer Global Research and Development, Groton Laboratories, Eastern Point Road, Groton Connecticut 06340

kevin.e.henegar@pfizer.com

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An operationally simple synthesis of N-BOC-2-hydroxymethylmorpholine (1) and N-BOC-morpholine-2-carboxylic acid (2) from epichlorohydrin has been developed. No chromatography is required in the processing, which allows high process throughput.

N-BOC-2-Hydroxymethylmorpholine (1) and the closely related *N*-BOC-morpholine-2-carboxylic acid (2) have been used extensively in the preparation of pharmacologically active compounds, both as the racemates and as single enantiomers. These include a number of CNS compounds such as reboxetine and related norepinephrine and norepinephrine-serotonin reuptake inhibitors (NRI and NSRI, respectively),¹ NRI and NSRI indazoles,² and other CNS structural classes.³ In addition, 1 and 2 have been used to prepare a wide variety of other pharmaceutically active compounds.⁴

Despite their widespread use, synthetic routes to these valuable compounds, either racemic or single enantiomer, are limited by expensive starting materials, processing issues, and lengthy and/or inefficient routes. For example, a 4-step synthesis of (*S*)-1 starting with (*S*)-3-amino-1,2-propanediol was recently reported as part of a synthesis of reboxetine.⁵ Although the starting material is commercially available, its high cost makes this route economically unattractive.⁶ In addition, the poor

solubility properties of the intermediates and the need for multiple chromatographies make scale-up of this synthesis problematic. Another short route uses the reaction of (S)benzylglycidyl ether with 2-aminoethyl hydrogen sulfate and base to generate the morpholine in a one-pot sequence.⁷ BOC protection and debenzylation yields 1. Unfortunately, tars are formed in this process which are difficult to remove. On a kilogram scale, the overall yield of 1 was 38%, but two distillations and three chromatographies were required to give material of acceptable quality, which seriously detracts from the chemical brevity of this synthesis. A more straightforward route also starting from (S)-benzylglycidyl ether has been described.⁸ This route has also been scaled but has seven isolated intermediates, three chromatographies, and a distillation, which limit throughput. A similar but even longer route starting with D-mannitol has also been described.9

Given the utility of **1** and **2**, there remains a need for a more efficient and scalable route to these valuable compounds. We describe herein a concise synthesis that can be used to make either enantiomer starting from inexpensive chiral epichlorohydrin. The basic synthetic approach is shown in Scheme 1.

^{*} Corresponding author.

 ^{(1) (}a) Fish, P. V.; Mackenny, M. C.; Stobie, A.; Wakenhut, F.; Whitlock,
 G. A. U.S. 2005/250775, Nov. 10, 2005; CAN 143:460166. (b) Fish, P. V.;
 Mackenny, M. C.; Stobie, A.; Wakenhut, F.; Whitlock, G. A WO 2005/105100,
 (Nov. 10, 2005; CAN 143:460165. (c) Barta, N. S.; Glase, S. A.; Gray, D. L.;
 Reichard, G. A.; Simons, L. J.; Xu, WU.S. 2005/245519, Nov. 3, 2005; CAN 143:440426.

⁽²⁾ Schelkun, R. M.; Yuen, P.-W. WO2006/056873, Jun. 1, 2006; CAN 145: 27985.

^{(3) (}a) Ono, S.; Yamafuji, T.; Chaki, H.; Maekawa, M.; Todo, Y.; Narita, H. EP383281, Aug. 8, 1994; CAN 114:81222. (b) Carruthers, N. I.; Gomez, L. A.; Jablonowski, J. A.; Keith, J. M.; Letavic, M. A.; Ly, K. S.; Miller, J. M. B.; Stocking, E. M.; Wolin, R. LWO2006/066197, Jun. 22, 2006; CAN 145:83244.
(c) Kojima, T.; Niigata, K.; Fujikura, T.; Tachikawa, S.; Nozaki, Y.; Kagami, S.; Takahashi, K. *Chem. Pharm. Bull.* 1985, *33*, 3766–74.

^{(4) (}a) Baldwin, J. J.; Claremon, D. A.; Tice, C.; Cacatian, S.; Dillard, L. W.; Ishchenko, A. V.; Yuan, J.; Xu, Z.; McGeehan, G.; Zhao, W.; Simpson, R. D.; Singh, S. B.; Flaherty, P. T.; Wery, J.-P. WO2006/042150, Apr. 20, 2006; CAN 144:432692. (b) Ko, S. S.; Delucca, G. V.; Duncia, J. V.; Santella, J. B., III; Wacker, D. A. WO 2000/035877, Dec. 17, 2000; CAN 133:58720. (c) Ackerman, J.; Banner, D.; Gubernator, K.; Hadvary, P.; Hilpert, K.; Mueller, K.; Labler, L.; Schmid, G.; Tschopp, T. EP468231, July 2, 1991; CAN 116:214908. (d) Amegadzie, A. K.; Beck, J. P.; Gardinier, K. M.; Hembre, E. J.; Ruble, J. C.; Savin, K. A.; Wakefield, B. DWO 2006/066174, June 22, 2006; CAN 145:83313. (e) Bower, J. F.; Poyser, J. P.; Turner, P.; Waterson, D.; Winter, J. WO2006/ 067401, June 29, 2006; CAN 145:103720. (f) Gudmundsson, K.; Miller, J. F; Turner, E. M. WO2006/020415, Feb. 23, 2006; CAN 144:233072. (g) Fink, B. E.; Gavai, A. V.; Vite, G. D.; Chen, P.; Mastalerz, H.; Norris, D. J.; Tokarski, J. S.; Zhao, Y.; Han, W.-C. WO2005/066176, July 21, 2005; CAN 143:153408. h) Gavai, A. V.; Han, W.-C.; Chen, P.; Ruediger, E. H.; Mastalerz, H.; Fink, B. E.; Norris, D. J. WO2005/065266, July 21, 2005; CAN 143:133402. (i) Kato, S.;
 Yamazaki, H.; Hirokawa, Y.; Suga, Y.; Yoshida, N.; Morikage, Y.; Morikage,
 K.; Oku, S. JP2004/277319, Oct. 7, 2004; CAN 141:307556. (j) Munson, M.; Mareska, D. A.; Kim, Y.; Groneberg, R. D.; Rizzi, J.; Rodriguez, M.; Kim, G.; Vigers, G.; Rao, C.; Balachari, D.; Harvey, D. US2004/192653, Sept. 30, 2004; CAN 141:314320. (k) Bhide, R.; Cai, Z.-w.; Qian, L.; Barbosa, S. WO2004/ 009784, Jan. 29, 2004; CAN 140:124558. (l) Luckhurst, C.; Perry, M.; Sanganee, H.; Springthorpe, B. WO2003/078395, Sept. 25, 2003; CAN 139:276907. (m) Mastalerz, H.; Zhang, G.; Tarrant, J. G.; Vite, G. D. WO2003/042172, May 22, 2003; CAN 138:401758. (n) Sanganee, H.; Springthorpe, B. WO2003/024962, Mar. 6, 2003; CAN 138:271690. (o) Gustafsson, J.; Hossain, N.; Nilsson, S. WO2003/018576, Mar. 6, 2003; CAN 138:221591. (p) Gustafsson, J.; Hossain, N.; Nilsson, S. WO2003/018566, Mar. 6, 2003; CAN138:221472. (q) Ko, S. S.; Delucca, G. V.; Duncia, J. V.; Santella, J. B., III; Wacker, D. A. WO2000/ 035877, June 22, 2000; CAN 133:58720. (r) Miyake, H.; Take, K.; Shigenaga, S.; Azami, H.; Sasaki, H.; Eikyu, Y.; Nakai, K.; Ishida, J.; Manabe, T.; Konishi, N.; Terasaka, T. WO98/57954, Dec. 22, 1998; CAN 130:81525. (s) Yanagisawa, H.; Fujimoto, K.; Shimoji, Y.; Kanezaki, T.; Iwata, N.; Kubo, Y. JP 04120021, 1992; CAN 117:251362. (t) Ackerman, J.; Banner, D.; Gubernator, K.; Hadvary, P.; Hilpert, K.; Mueller, K.; Labler, L.; Schmid, G.; Tschopp, T. EP468231, Jan. 29, 1992; CAN 116:214908. (u) Nilsson, M.; Zhou, X.-X.; Oden, L.; Classon, B.; Noren, R.; Grabowska, U.; Jackson, P.; Fallon, P.; Carr, A.; Liley, M.; Tozer, M.; Johnson, T.; Diaz, V.; Crespo, L.; Kangasmetsa, J.; Bonnaud, T. WO2005/ 066180, July 21,2005; CAN 143:153706.

⁽⁵⁾ Brenner, E.; Baldwin, R. M.; Tamagnan, G. Org. Lett. 2005, 7, 937-939.

⁽⁶⁾ The lowest bulk quote we received for (S)-3-amino-1,2-propanediol was 10 kg at \$6000/kg).

⁽⁷⁾ Ackerman, J.; Banner, D.; Gubernator, K.; Hadvary, P.; Hilpert, K.; Mueller, K.; Labler, L.; Schmid, G.; Tschopp, T. EP468231, Jan 29, 1992; CAN 116:214908.

⁽⁸⁾ Berg, S.; Larsson, L.-G.; Renyi, L.; Ross, S. B.; Thorberg, S.-O.; Thorell-Svantesson, G. J. Med. Chem. 1998, 41, 1934–1942.

⁽⁹⁾ Yanagisawa, H.; Kanazaki, T. Heterocycles 1993, 35, 105-9.

SCHEME 1



Reaction of chiral epichlorohydrin 3^{10} with a protected ethanolamine 4 would yield the chlorohydrin 5. Epoxide formation (6) from the chlorohydrin was expected to be facile, and then further reaction of the epoxide would induce cyclization to the morpholine-oxazepine mixture (7 and 8).

We are aware of a single example of this type of basepromoted cyclization in the literature, which claimed a 9:1 ratio of morpholine to oxazepine for the *N*-methyl compounds derived from racemic epichlorohydrin and *N*-methylethanolamine (Scheme 1, $R = CH_3$). These results are hard to interpret since the ratio appears to be reported for a purified sample and the actual yield is unclear.¹¹

N-Benzylethanolamine **9** is also known to react with racemic epichlorohydrin to give the racemic chlorohydrin **10**, which has been cyclized to give **11** (Scheme 2). The cyclization was done in concentrated sulfuric acid at 95–150 °C and the stereochemical outcome of this cyclization is unknown.¹²

N-Benzylethanolamine (9) was chosen for this process since removal of the benzyl group should be facile and the benzyl group provides a convenient chromophore for reaction monitoring. Reaction of N-benzylethanolamine with (S)-epichlorohydrin 3 cleanly yielded the chlorohydrin 12 (Scheme 3). In contrast to the reported reaction with racemic epichlorohydrin,¹² which used a large excess of epichlorohydrin, we ran the reaction with a 1:1 molar ratio of N-benzylethanolamine:epichlorohydrin to minimize consumption of the chiral epichlorohydrin. This reaction proceeded only in protic solvents and gave essentially no product in solvents such as DMF, THF, CH₃CN, or toluene. The reaction ran well in alcohols such as methanol or IPA and was substantially accelerated by addition of water. In addition, reaction in aqueous alcohols minimized impurity formation. The chlorohydrin was relatively unstable; isolated samples lost HPLC potency and degraded so it is not normally isolated.

Conversion of the chlorohydrin 12 to the epoxide 13 was facile. Epoxide 13 underwent further reaction readily and for

SCHEME 3



isolation the epoxide, formation was best done under phase transfer conditions. The epoxide was also relatively unstable and was not normally isolated; purified samples cyclized in a few months when stored in a refrigerator and faster at ambient conditions.

A survey of Lewis acid conditions for the conversion of **13** to **14** and **15** showed slow and very messy reaction with LiOTf in CH₃CN or *t*-BuOH, no reaction with LiClO₄ in CH₃CN, *t*-BuOH, or Et₂O, very messy reaction with trifluoroacetic acid, and no reaction with methanesulfonic acid. Exposure of the epoxide to lanthanide triflates (La(OTf)₃, Yb(OTf)₃, Nd(OTf)₃) in 2-propanol or CH₃CN gave no reaction.

Cyclization of **13** to **14** and **15** was promoted by hydroxide. Concentrated NaOH or KOH worked but the reactions were nonhomogeneous and became unstirrable due to salt precipitation. LiOH was ineffective and CsOH also worked but was no different than NaOH or KOH. Aqueous tetraalkylammonium hydroxide solutions gave mixtures that were homogeneous throughout the reaction. No difference was seen in the ratio of **14** to **15** with various hydroxides (Me₄NOH, Et₄NOH, *n*-Pr₄NOH, *n*-Bu₄NOH, and BnNMe₃NOH were examined). The ratio of **14** to **15** was invariably about 70:30. There was minimal temperature effect on this ratio and the reaction was run at about 20–25 °C. *n*-Bu₄NOH was originally used but extraction of the product was inefficient and there was carryover of tetrabutylammonium residues into the organic phase. The more hydrophilic Et₄NOH was chosen to overcome these issues.

Stereochemical correlation was done starting with (R)epichlorohydrin **3**. This yielded the (S)-enantiomer of **1** with no detectable enantiomer by chiral HPLC analysis, demonstrating that the epoxide opening proceeded with complete inversion. However, when chlorohydrin **12** was used that had been held as a concentrated oil for about a week before use, the **1** obtained had about 15% of the wrong enantiomer. Racemization is presumably proceeding through intermediacy of the azetidinium

^{(10) (}*R*)-Epichlorohydrin used for this study was purchased for 470/kg in 5 kg lots. Both enantiomers of epichlorohydrin are available in bulk for 470/kg.

 ⁽¹¹⁾ Buriks, R. S.; Lovett, E. G. J. Org. Chem. 1987, 52, 5247–5254.
 (12) (a) Loftus, F. Synth. Commun. 1980, 10, 59–73. (b) Lainton, J. A. H.;

^{(12) (}a) Loftus, F. Synth. Commun. **1980**, 10, 59–73. (b) Lainton, J. A. H.; Allen, M. C.; Burton, M.; Cameron, S.; Edwards, T. R. G.; Harden, G.; Hogg, R.; Leung, W.; Miller, S.; Morrish, J. J.; Rooke, S. M.; Wendt, B. J. Comb. Chem. **2003**, 5, 400–407.

SCHEME 4



species.^{13,14} Reaction starting with (S)-epichlorohydrin predictably yielded the (R)-enantiomer of **1**.

Stereochemical analysis of the oxazepine 15 derived from (R)-epichlorohydrin was done. Chiral HPLC assay showed it to be essentially a single enantiomer, indicating it is formed by a competing 7-endo cyclization and not via an azetidinium species, which would have given racemic oxazepine product.

In the final optimized conditions, chlorohydrin formation, epoxidation, and cyclization were run in one pot without isolation of the intermediates **12** and **13**. The reaction to form **12** was done in 1:1 2-propanol-water at 20–25 °C. When this reaction was done, 1.2 equiv of 35% aqueous Et₄NOH was added. Formation of epoxide **13** was rapid and cyclization to **14** and **15** required about 2 h.

Selective succinoylation¹⁵ of the morpholine primary alcohol to hemisuccinate 16 allowed nonchromatographic separation of the mixture of 14 and 15. Selectivity for the primary vs secondary alcohol was about 6:1 at rt but increased at lower temperatures. Reaction at -25 °C gave rapid reaction with the primary alcohol and good selectivity. Simple distillation of the crude mixture of 14 and 15 to remove high-boiling impurities gave better control over succinic anhydride stoichiometry and gave cleaner final product. Succinovlation was fast and selective in CH₂Cl₂ with triethylamine or other nonhindered bases such as dimethylethylamine or N-methylpyrrolidine, and was sluggish with diisopropylethylamine as the base or in ethyl acetate or methyl tert-butyl ether. One equivalent of succinic anhydride based on the morpholine HPLC area percent of 14 was added in 2 or 3 portions. After 30 min, dilute aqueous ammonia was added to extract the hemisuccinate into the aqueous phase. Use of aqueous Na₂CO₃ or K₂CO₃ for the extractions led to serious emulsions. The succinate 16 was readily hydrolyzed by addition of NaOH. The isolated purified 14 contained <1% oxazepine 15. Hydrogenation and BOC protection yielded 1 with >99%ee in 41% overall yield from (R)-epichlorohydrin.

Conversion of this mixture to the carboxylic acid (2) was straightforward (Scheme 4). The undistilled mixture of 14 and

15 was hydrogenated to remove the benzyl groups and the nitrogens protected to give the BOC derivatives 17 and 18. Tempo oxidation converted the primary alcohol of the morpholine 14 to the carboxylic acid 2 and the secondary alcohol of 15 to the ketone 19. These products were easily separated by simple acid-base extractions. Overall yield of 2 from epichlorohydrin was 39%, with no chromatography or distillation. The product assayed at >99% ee by chiral HPLC.

Experimental Section

(S)-N-BOC-2-Hvdroxymethylmorpholine (1). N-Benzylethanolamine 9 (151.2 g, 1.0 mol), water (100 mL), and 2-propanol (100 mL) were charged to a jacketed 2 L reactor with a 20 °C circulating jacket. (R)-Epichlorohydrin 3 (97.15 g, 1.05 mol) was added and the solution was stirred overnight. A 35 wt % aqueous solution of Et₄NOH (495 mL, 1.3 mol) was added over <5 min. The mixture exothermed to +29 °C in about 30 min, and then the temperature returned to ambient. After 4 h, the reaction was quenched with 1 M HCl (150 mL, final pH: 9). Water (350 mL) was added and the mixture was extracted with CH_2Cl_2 (3 \times 500 mL). The extracts were combined and evaporated to an oil, which was distilled through a short path under vacuum (0.5 mm, bath 175-190 °C, bp 150-180 °C) to yield a clear colorless distillate (169.9 g) and about 20 g of clear amber still bottoms. The distillate was 67.7:32.3 14:15 by HPLC; the still bottoms were a gross mixture containing little 14.

The distillate (**14** and **15**, 169.9 g) and triethylamine (115 mL, 0.81 mol) were dissolved in 1500 mL of CH₂Cl₂ and cooled to -25 °C (± 3 °C). Succinic anhydride (49.0 g, 0.49 mol) was added in two portions over 45 min and then stirred for 2 h. Dilute aqueous ammonia (40 mL of 28% diluted to 440 mL volume) was added and the mixture was warmed to rt and stirred for 30 min. The organic phase was extracted with dilute aqueous ammonia (2×40 mL of 28% diluted to 440 mL volume). The combined aqueous phases were extracted with CH₂Cl₂ (1×400 mL). The aqueous phase was basified with 50% NaOH (40 mL) and stirred overnight. The solution was extracted with CH₂Cl₂ (3×400 mL), dried briefly over Na₂SO₄, filtered, and evaporated to yield 91.2 g (44% overall) of **14** as a colorless oil.

The product was hydrogenated over Pd/C (50 psi, 10% Pd/C, 50% water wet) in methanol. The solution in methanol (750 mL volume) after catalyst filtration was stirred at rt and BOC₂O (96 g, 0.44 mol) in 100 mL of methanol was added in portions over 90 min. The solution was stirred for 30 min after completion of the addition. An additional 5 g of BOC₂O dissolved in 10 mL of methanol was added and the solution was stirred for 30 min. N,N-Dimethylethylenediamine (66 mmol, 7.2 mL) was added and the mixture was stirred for 30 min. The solution was concentrated on a rotovap to an oil. The oil was partitioned between CH₂Cl₂ (1L) and 100 mL of 1 M HCl and 100 mL of water. The CH₂Cl₂ phase was extracted again with 100 mL of 1 M HCl and 100 mL of water. The combined aqueous phases were back extracted with 500 mL of CH₂Cl₂. The combined CH₂Cl₂ solutions were washed with 200 mL of 1 M Na₂CO₃. The CH₂Cl₂ solution was evaporated to an oil and then heptane $(2 \times 1 L)$ was added and evaporated. The resulting oil solidified on standing to yield a white solid 1 (90.2 g, 41.8% overall from (R)-epichlorohydrin): mp 61-64 °C (lit.⁵ mp 60-62 °C); $[\alpha]^{25.3}_{D} + 18.97^{\circ}$ (c 0.2, CHCl₃); (lit.⁵ $[\alpha]^{20}_{D} + 20.7$ (c 1.01, CHCl₃)); ¹H NMR (399.76 MHz, CDCl₃) & 1.43 (s, 9H), 2.19 (br s, 1H), 2.64–2.79 (m, 1H), 2.82–2.99 (m, 1H), 3.42–3.60 (m, 3H), 3.60-3.67 (m, 1H), 3.73-3.94 (m, 3H); ¹³C NMR (100.53 MHz, CDCl₃) & 28.5, 42.5–46.1 (br), 63.6, 66.5, 76.0, 80.3, 154.9; Chiral HPLC, Chirapak AD 4.6 mm \times 250 mm, 10 μ m, 92:8:0.1 hexane: IPA:TFA, 1.0 mL/min, 215 nm; S, 8.7 min; R, 9.8 min; ROI < 0.1%. Anal. Calcd for $C_{10}H_{19}NO_4$: C, 55.28; H, 8.81; N, 6.45. Found: C, 55.43; H, 8.90; N, 6.44.

(S)-2,4-Morpholinedicarboxylic Acid, 4-(1,1-Dimethylethyl) Ester 2. N-Benzylethanolamine 9 (12.6 g, 83.3 mmol) and (R)-

^{(13) (}a) Gaertner, V. R. *Tetrahedron Lett.* **1964**, *3*, 141–146. (b) Gaertner,
V. R. *J. Org. Chem.* **1967**, *32*, 2972–2976. (c) Gaertner, V. R. *J. Org. Chem.* **1968**, *33*, 523–530. (d) Oh, C. H.; Rhim, C. Y.; You, C. H.; Cho, J. R. Synth. Commun. **2003**, *33*, 4297–4302.

⁽¹⁴⁾ The HPLC purity of the isolated chlorohydrin had dropped from about 90 area % to less than 60% during storage.

⁽¹⁵⁾ Formation of the hemisuccinate 16 was confirmed by mass spectral data. Other anhydrides examined either gave sluggish reaction and poor selectivity (phthalic anhydride, glutaric anhydride) or poor partitioning during the aqueous base extractions (hexahydrophthalic anhydride).

epichlorohydrin **3** (8.09 g, 87.5 mmol) were stirred in a 35 °C oil bath with 12 mL of 2-propanol. The solution was cooled to rt. Et₄NOH (35% aqueous; 43 mL, 99 mmol) was added and the mixture was allowed to stir overnight at toom temperature. The pH was adjusted to 9–10 with concd HCl. Water (50 mL) and MTBE (50 mL) were added. The phases separated and the aqueous phase was extracted with 50 mL of methyl *tert*-butyl ether. The combined organic phases were dried over Na₂SO₄, filtered, and evaporated to yield the crude product (**14** and **15**, 15.28 g). The material was hydrogenated in methanol with Pd/C (50 psi, 10% Pd/C, 50% water wet). Yield 8.16 g of a slightly yellow oil.

The oil was stirred with THF (100 mL). A small amount of white solids formed which were removed by filtration. BOC₂O (69 mmol, 14.6 g) was added in portions as a solid. The solution was stirred overnight a rt and then evaporated to an oil. Methyl *tert*-butyl ether (2 × 100 mL) was added and evaporated to yield a mixture of **17** and **18**. The oil was dissolved in CH₂Cl₂ (67 mL). Water (33 mL), Bu₄NBr (0.93 g, 2.9 mmol), and 4-acetamidotempo (0.06 g, 0.28 mmol) were added and the two-phase mixture was stirred at 0 °C. A solution of 70 mL of 12.5% NaOCI and 0.3 g of NaHCO₃ and 20 mL of water was added over 1 h. The excess NaOCI was quenched with sodium sulfite and the pH (about 7) was adjusted to >12 with 50% NaOH. The phases were separated and the aqueous phase was extracted with 20 mL of CH₂Cl₂. The pH of

the aqueous phase was adjusted to <2 with concd HCl and the solution was extracted with CH₂Cl₂ (3 × 15 mL). The organic solutions were combined, dried over Na₂SO₄, and evaporated to yield **2** as a white solid (7.65 g, 39% overall): mp 127–134 °C dec; $[\alpha]^{20}_{\rm D}$ +30.58 (*c* 0.5, EtOH); ¹H NMR (399.76 MHz, CDCl₃) δ 1.42 (s, 9H), 2.99–3.15 (m, 2H), 3.52–3.60 (m, 1H), 3.71–3.80 (m, 1H), 3.93–4.02 (m, 1H), 4.04–4.17 (m, 2H), 6.83 (br s, 1H); ¹³C NMR (100.53 MHz, CDCl₃) δ 28.5, 43.0 (br), 45.6 (br), 66.2, 73.7, 81.2, 154.8, 172.2; low resolution mass spec, nominal *m/e* calcd for C₁₇H₂₁O₄ (M + H)⁺ 289, found *m/z* 289 [M +1]⁺ (Ion Mode: APCI); Chiral HPLC, Chirapak AD 4.6 × 250 mm, 10 μ m; 95:5:0.1 hexane:ethanol:TFA; 0.5 mL/min; 215nm; *R*, 29.8 min; *S*, 27.6 min; ROI < 0.1%. Anal. Calcd for C₁₀H₁₇NO₅: C, 51.94; H, 7.41; N, 6.06. Found: C, 51.95; H, 7.40; N, 6.01.

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Supporting Information Available: Experimental procedures and spectral data for intermediates. This material is available free of charge via the Internet at http://pubs.acs.org.

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